

# Biomarker Data 101

## OR

# Bridging the Transdisciplinary Communication Gap

Judy-Anne Chapman, Ph.D., P.Stat., PStat<sup>®</sup> (ASA)



Information about SSC Accreditation  
of Professional Statisticians at  
<http://www.ssc.ca/en/accreditation>

# Overview of Talk

1. What is a biomarker?
2. Biomarker use.
3. Breast cancer biomarker assessments.
4. Sources of biomarker assay variability.
5. Case studies.
6. Work in progress.

# What is a biomarker?

## **NIH (Environmental Health Sciences):**

“Biomarkers are key molecular or cellular events that link a specific environmental exposure to a health outcome.”

## **NCI:**

“A biological molecule found in blood, other body fluids, or tissues that is a sign of a normal or abnormal process, or of a condition or disease.”

# Biomarker Use

1. Risk of developing disease.
2. Diagnosis.
3. Response to therapy: success, toxicity.
4. Risk of sequelae: same/other disease, death.

**→ Tension/tradeoff for actionable targets/therapy**

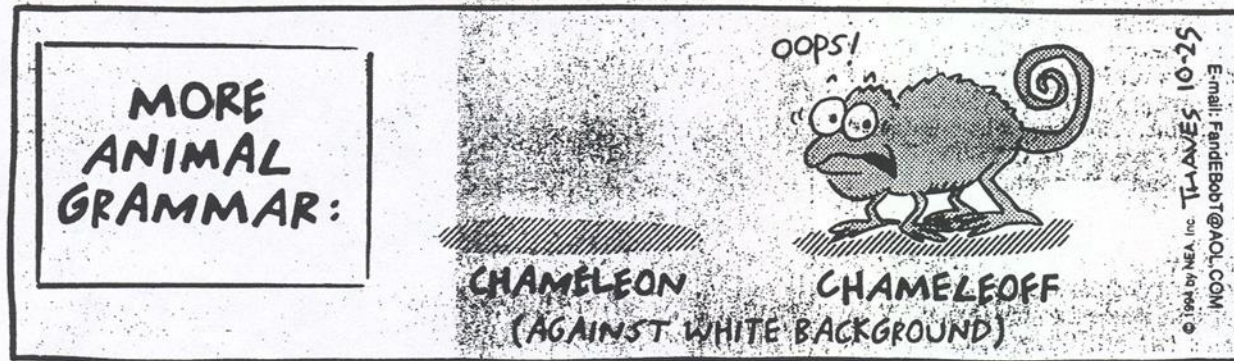
Biomarker data (mostly measured, continuous)

vs

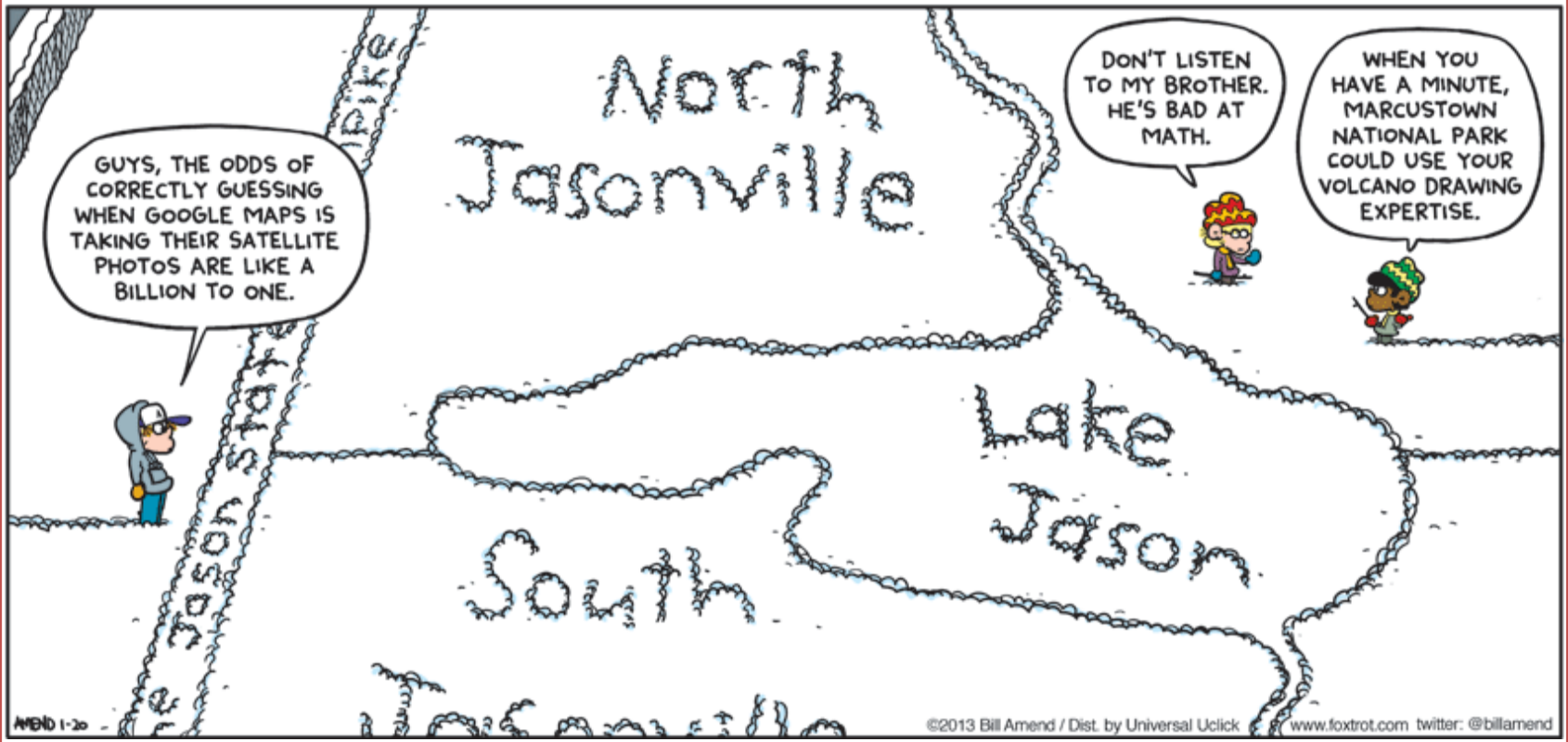
Frequently-assessed for action (categorical).

# DECISION THEORY

Frank and Earnest



# Chance of hitting the right cut-point(s) for clinical action?



# Breast Cancer Biomarker Assessments

## 1. Determinant of endocrine therapy:

Up to 20% of immunohistochemical (IHC) Estrogen Receptor (ER) and Progesterone Receptor (PR) inaccurate:

ASCO/CAP IHC Guidelines (Hammond MEH, et al. J Clin Oncol 2010)

- ER+, PR+ with  $\geq 1\%$  cells staining;
- Establishment of more external quality assurance programs.

## 2. Determinant of anti-HER2 therapy:

Up to 20% of human epidermal growth factor receptor 2 (HER2) gene inaccurate:

ASCO/CAP Guidelines (Wolff AC, et al. J Clin Oncol 2007; 2012)

- Establishment of cut-points;
- Establishment of more external quality assurance programs.

## 3. Ki67 Working Group:

NIH moratorium on using banked Breast Cancer trial specimens to assess ki67: (Dowsett M, et al. J Natl Cancer Inst 2011)

- Work in progress (Lisa McShane involved as statistician)

# Sources of Biomarker Assay Variability

## **Kananaskis Working Group:**

Assessing genetic markers of tumour progression in the context of intratumour heterogeneity.

Chapman JW, Wolman E, Wolman SR,...

Shankey TV. Cytometry 1998; 31:67-73.

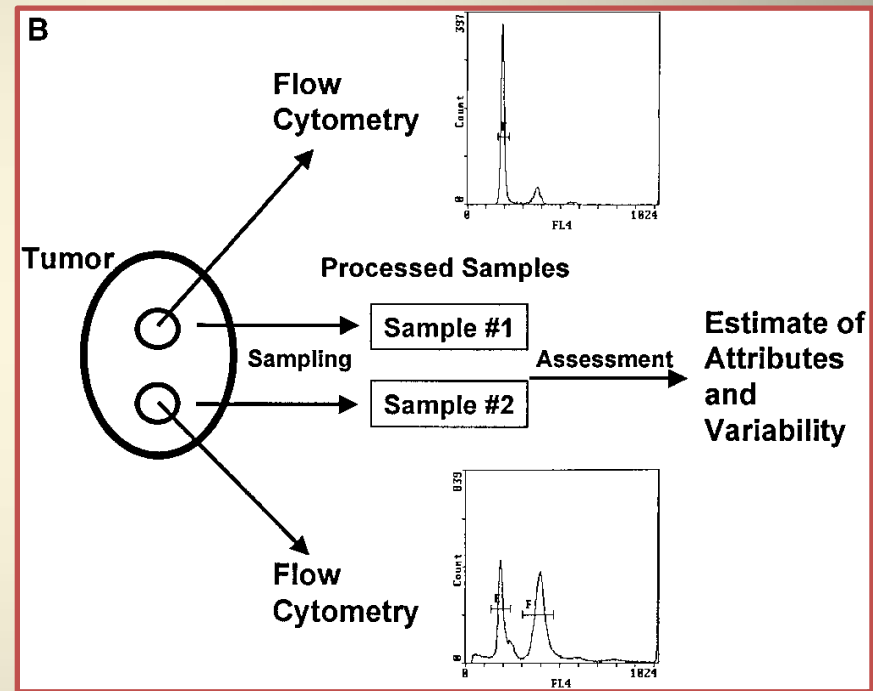


# Quantification of Heterogeneity

Assessed tumour heterogeneity encompasses:

1. Reproducibility error;
2. Intratumour heterogeneity, which may change in a tumour with time
  - Not routinely assessed;
3. Intertumour differences among tumours.

# Demonstrated Tumour Heterogeneity

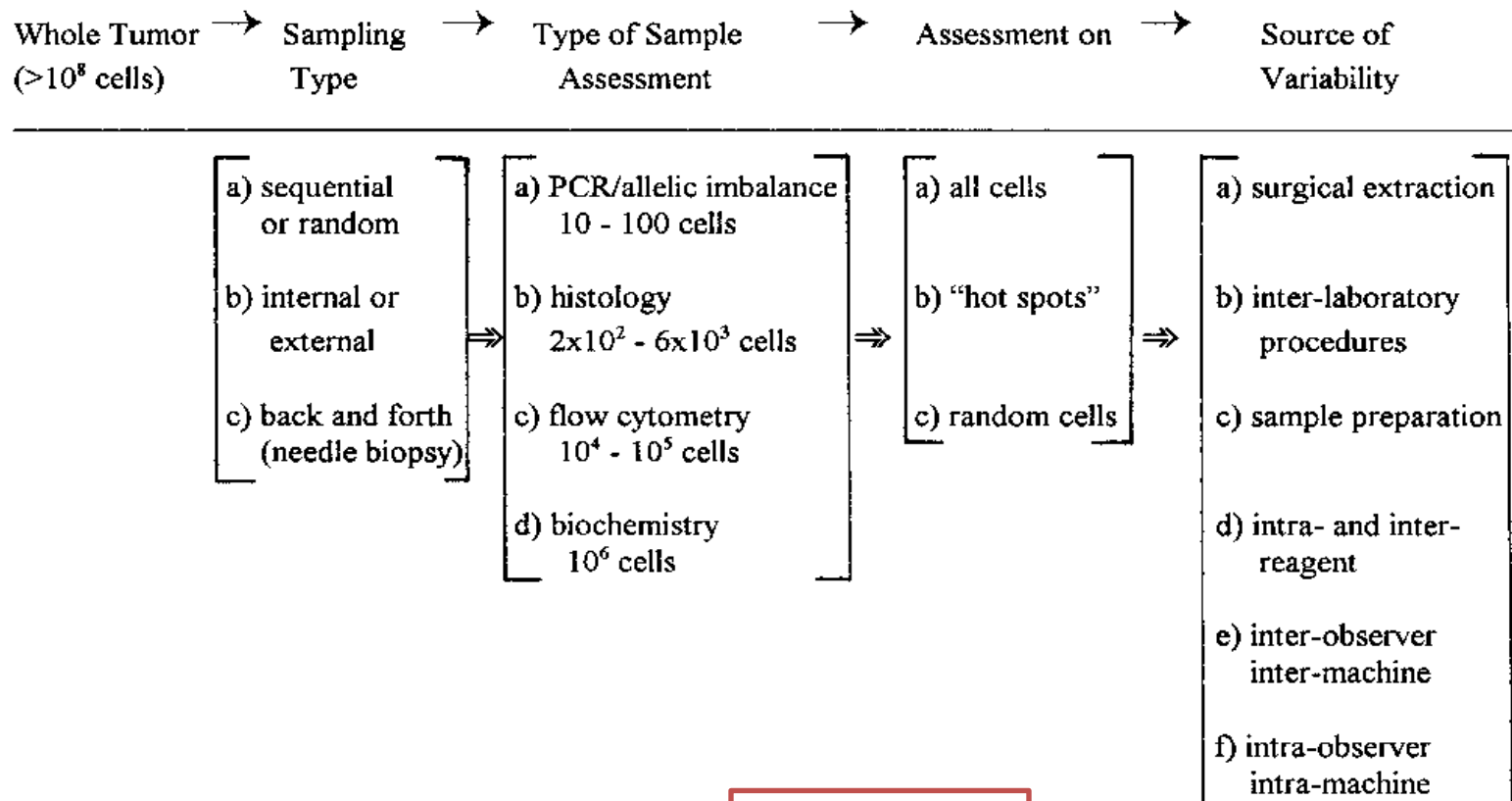


Courtesy of Vince Shankey [Chapman JW,...,Shankey TV. Cytometry 31:67–73 (1998)]

# Underlying Tumour Heterogeneity:

Chapman, et al [Cytometry 31:67–73 (1998)]

## C Sources of Variability



# Pragmatic Handling of Different Methods, Sample Sizes (# of cells), Variability

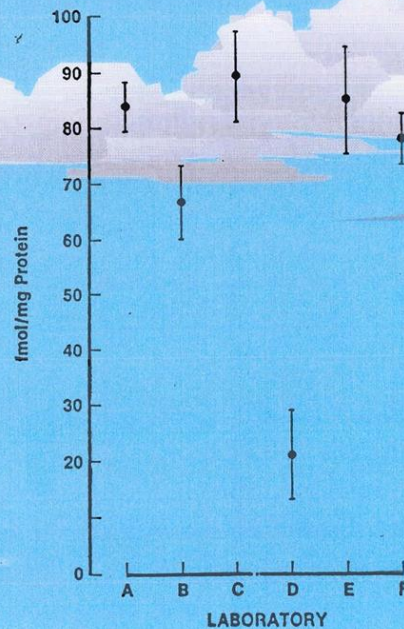
- Currently, assume + is +, different assays.
- Assume measuring similar process(es).
- Expect differences in location/scale.
- Where possible, investigate using multivariable, continuous biomarker data:  
(Kananaskis Working Group, Cytometry 1998;

REMARK 2012, BMC Medicine 2012, 10:51  
Altman DG, McShane LM, Sauerbrei W,  
Taube SE)

# Case Study: Biochemical Dextran-Coated Charcoal (DCC; Radioligand binding) Assay for ER

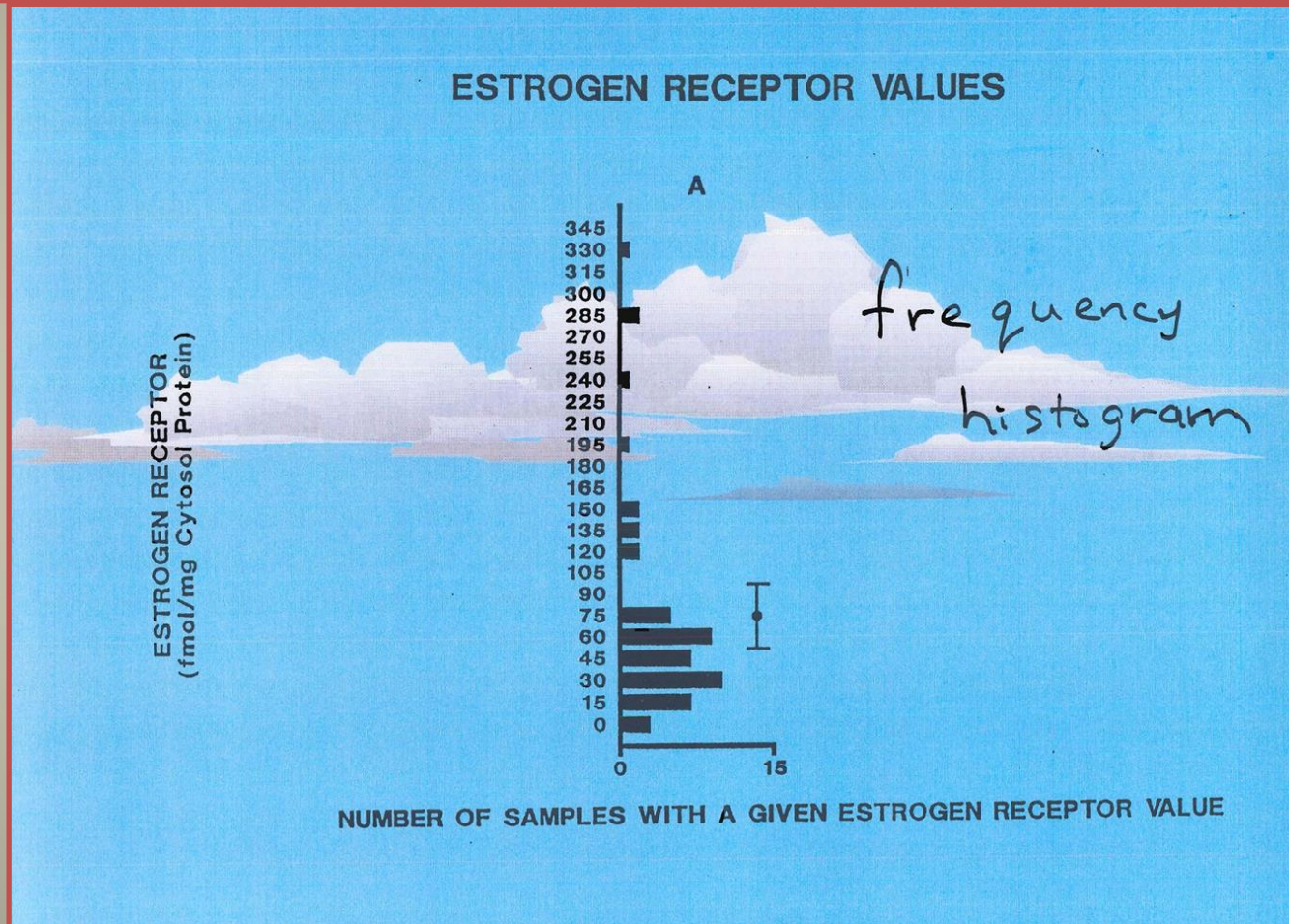
## 5. Hormone Receptor Status

ER FOR LABORATORIES USING THE SAME POWDERS, VARIOUS METHODOLOGIES



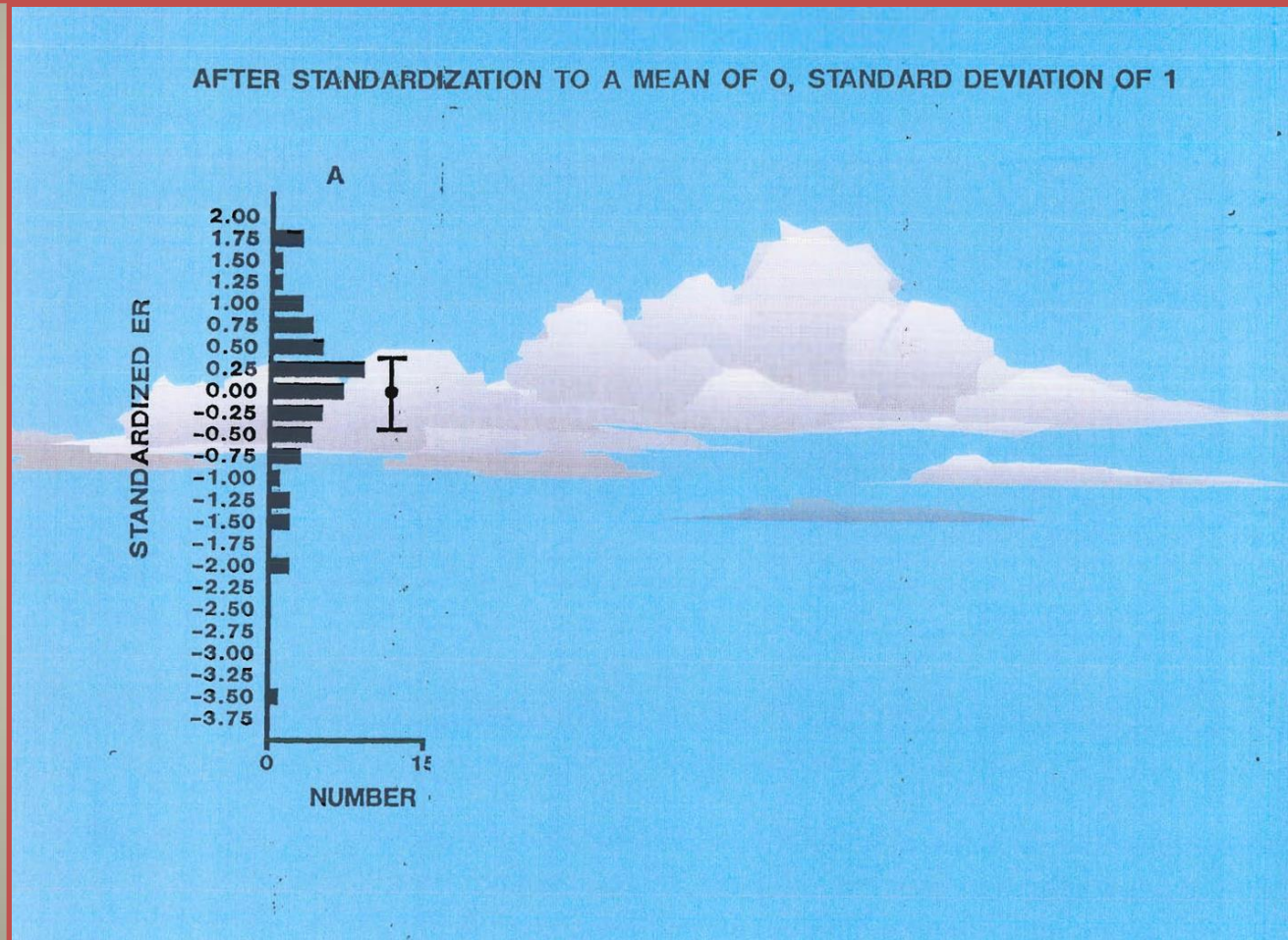
Ryan, et al. (1985)

# ER Frequency Histogram: Laboratory A



Chapman JW, Mobbs BG, et al. J Steroid Biochem Molec Biol 1993; 45:367-373.

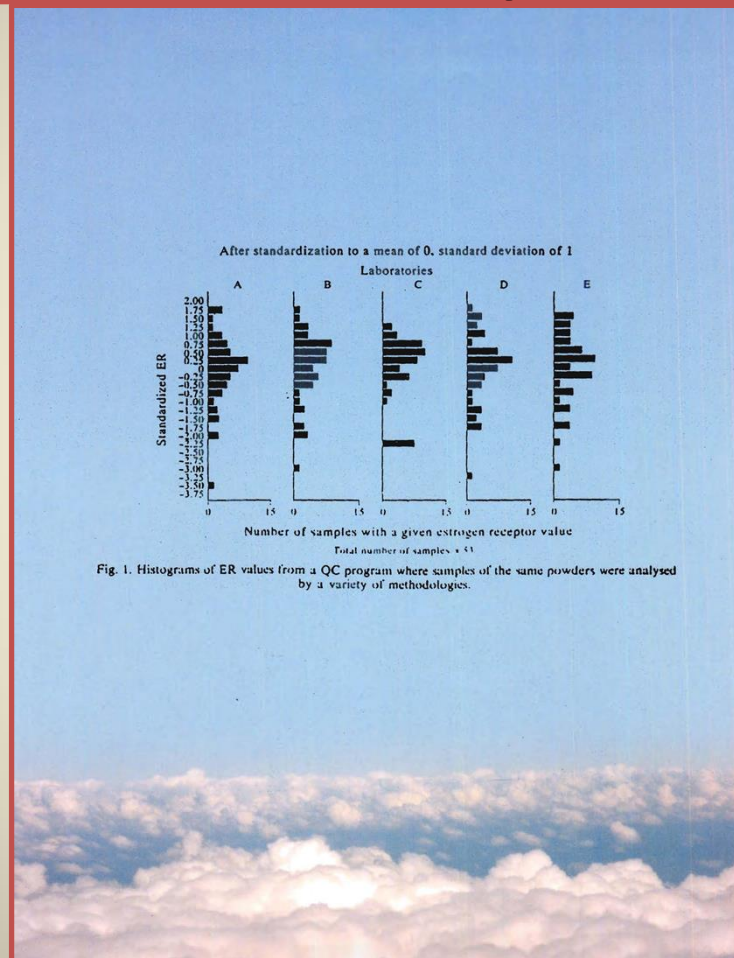
# Statistically Standardized Logarithm (ER) for Laboratory A



Chapman JW, Mobbs BG, et al. J Steroid Biochem Molec Biol 1993; 45:367-373.

JW Chapman

# Comparison of Statistically Standardized ER by Laboratory

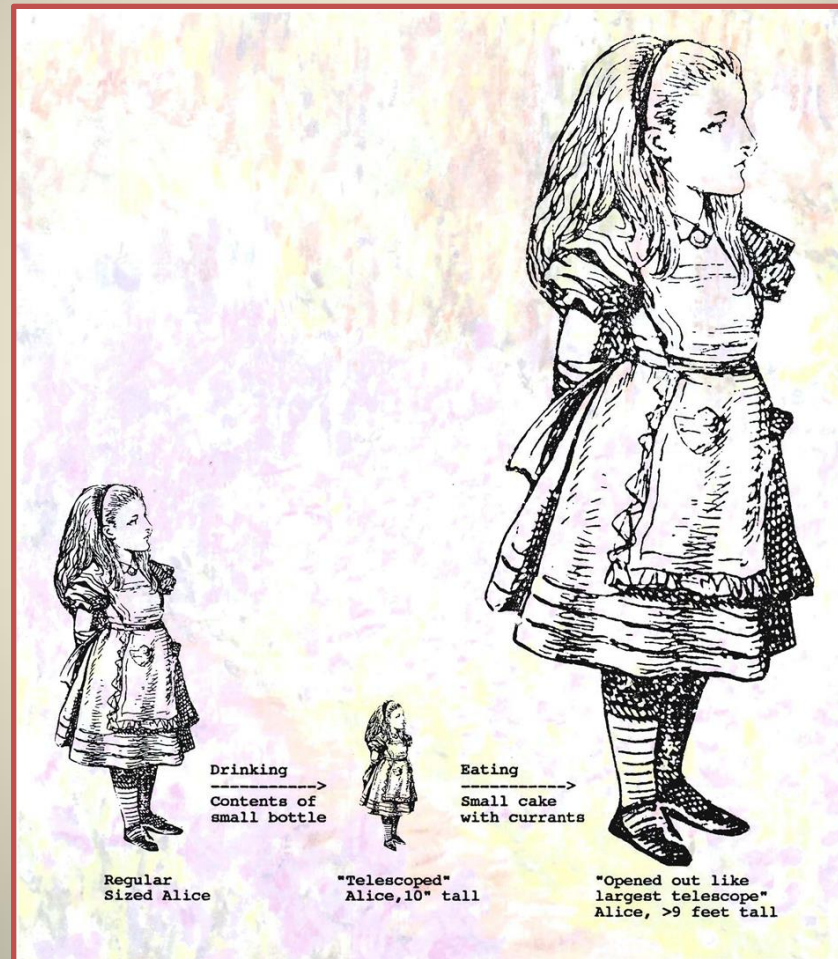


Chapman JW, Mobbs BG, et al. J Steroid Biochem Molec Biol 1993; 45:367-373.

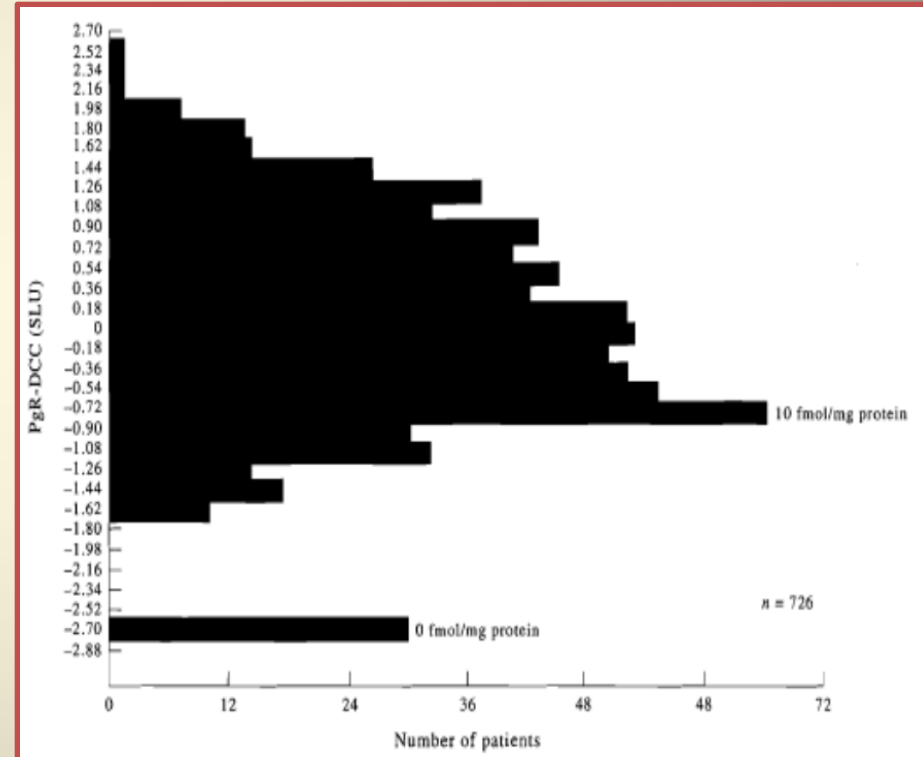
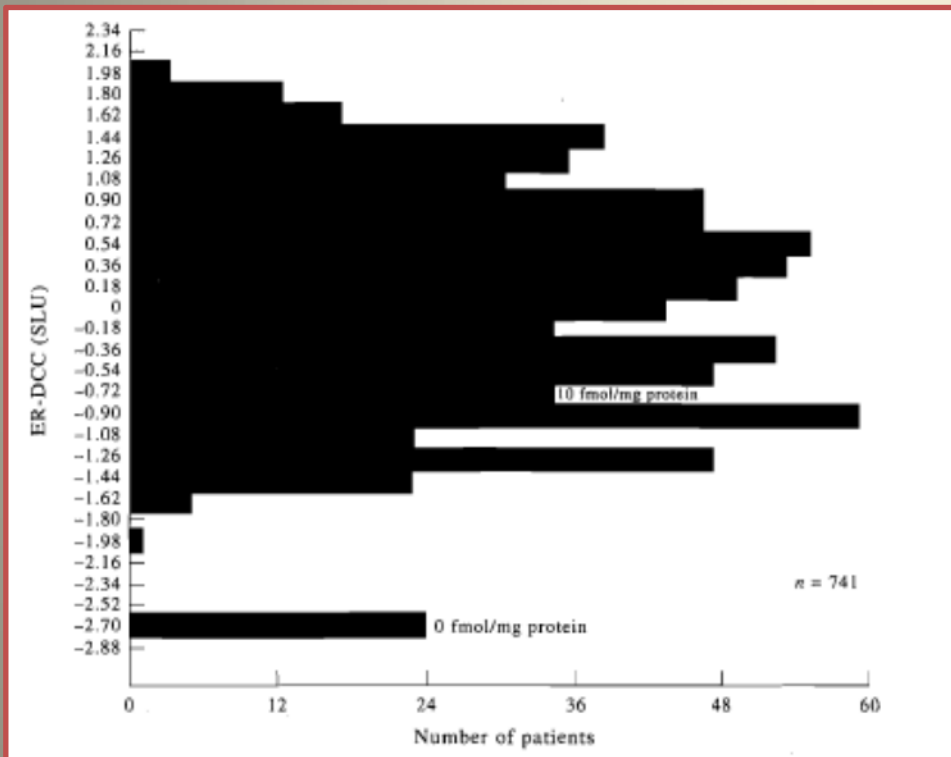
JW Chapman



# Double 1 to 1 Transformation

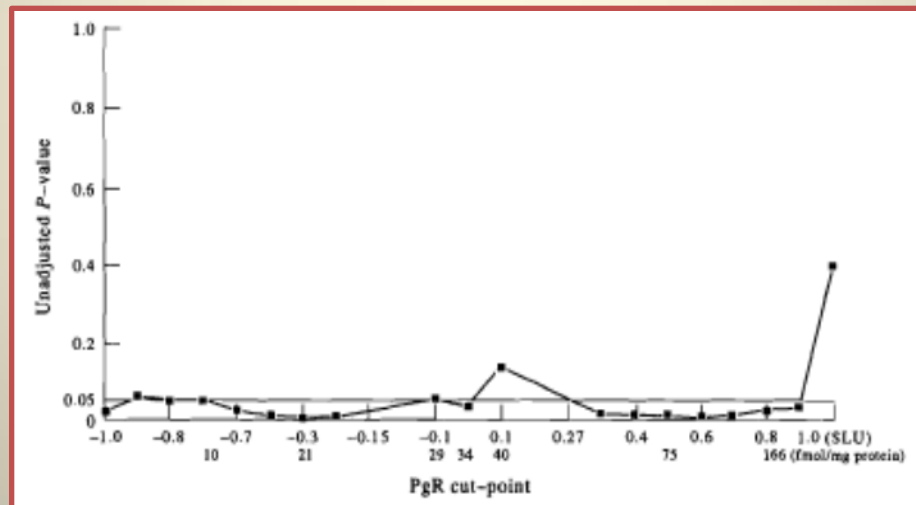
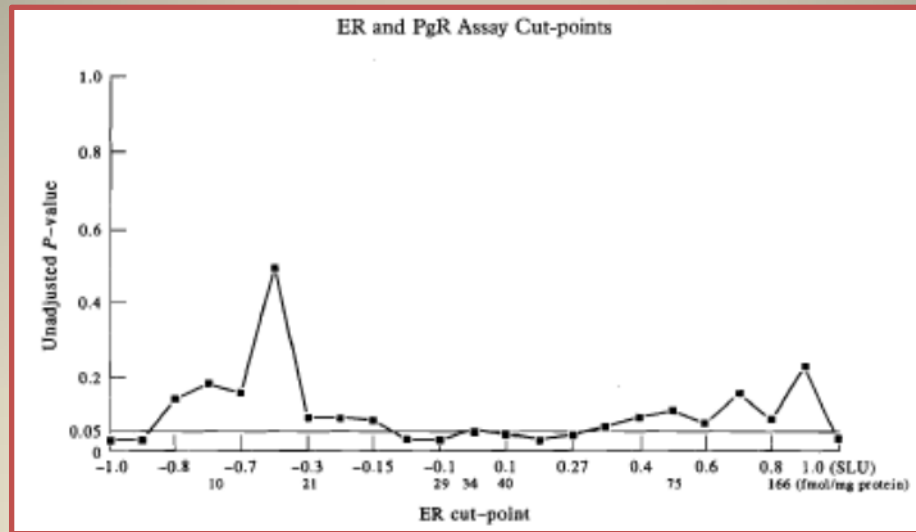


# Frequency Histograms of ER-DCC in Standardized Log Units (SLU)



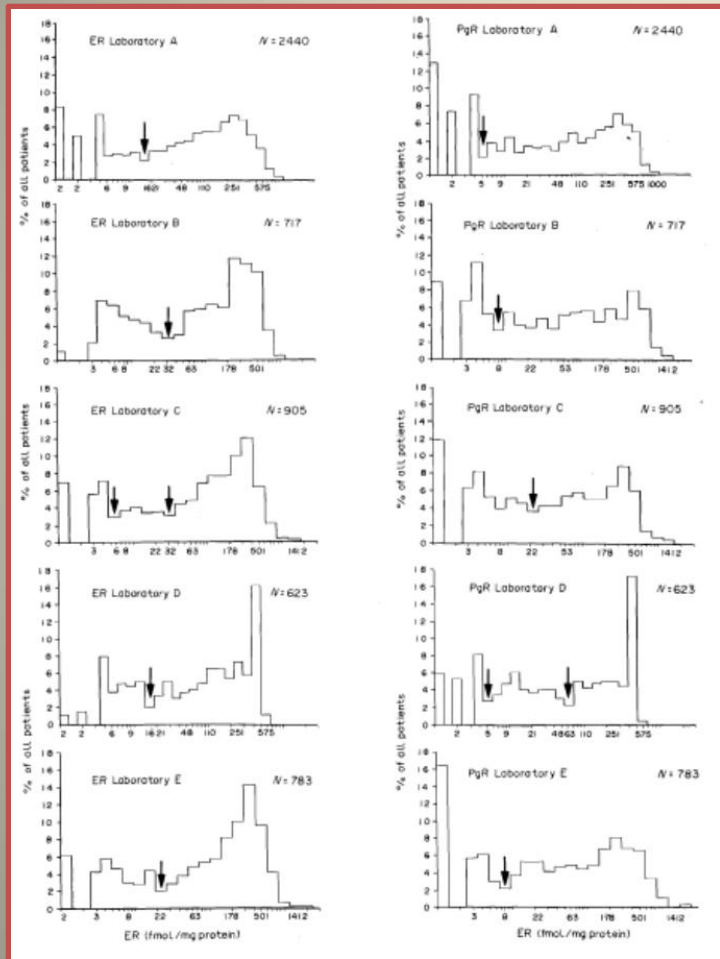
Chapman JW, Mobbs BG, et al, Eur J Cancer, 1996)

JW Chapman



Chapman JW, Mobbs BG, et al. J Steroid Biochem Molec Biol 1996; 57:323-328.

# Biochemical ER and PR by Enzymeimmunoassay (EIA; double monoclonal antibody)



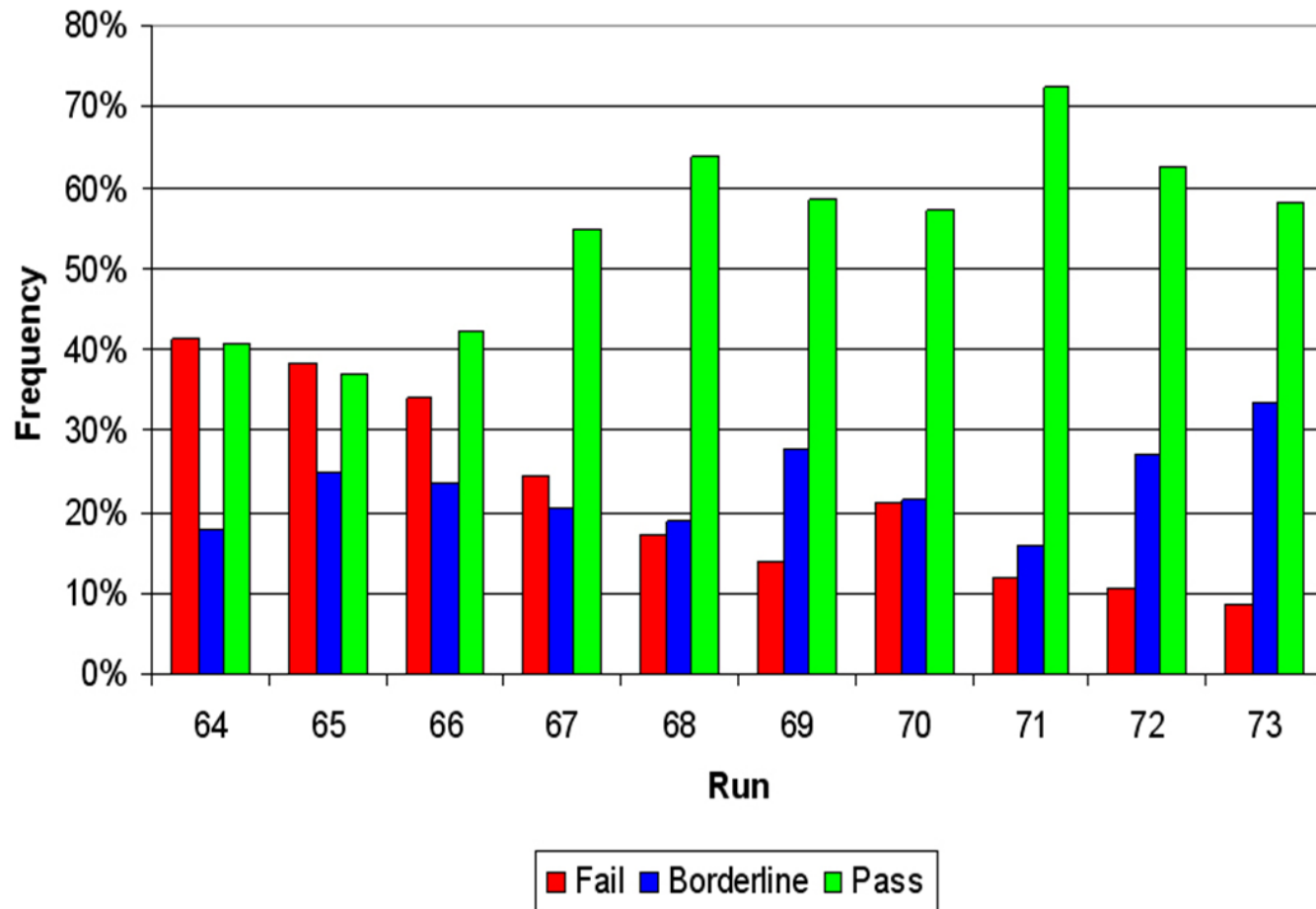
One conclusion in paper, about bimodality:  
“It is conceivable that oestradiol-binding mutant ERs may occur which have altered binding properties for the antibodies used In the EIA.”  
Postulated existence of ER $\beta$

Mobbs BG, Chapman JW, et al. Eur J Cancer 1993; 29:1292-1297

JW Chapman

# IHC Estrogen Receptor: UK NEQAS ICC & ISH

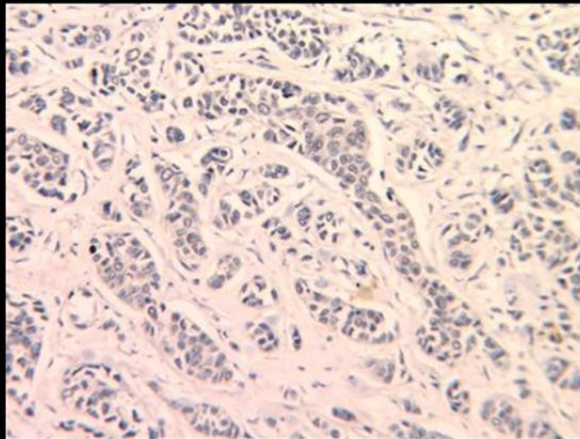
Courtesy of B Jasani, M Ibrahim, K Miller



# Example of Order of Difference

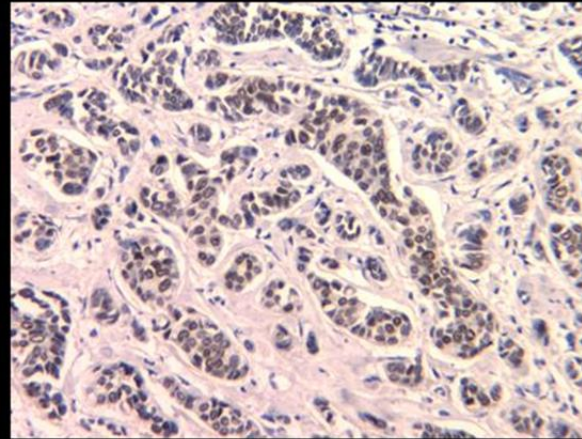
## ER: In-house Staining Can Sometimes Cause Concern

ER: Stained by Participant



Diagnosis: Negative

Same ER: Stained by NEQAS

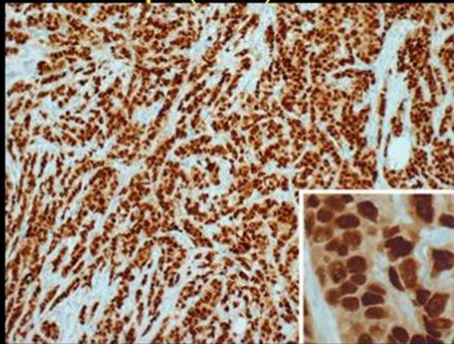


Diagnosis: >50% +

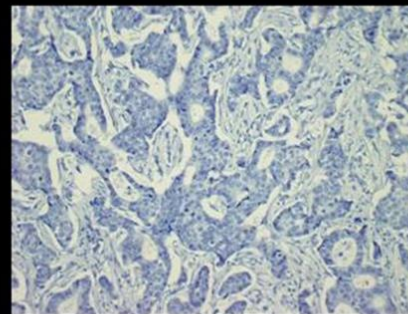
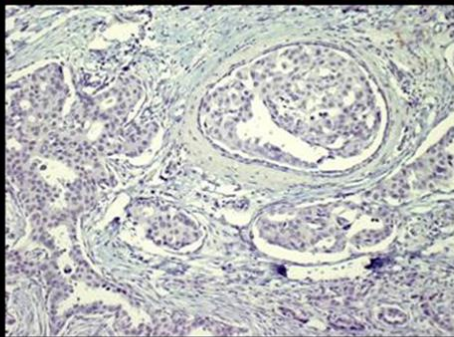
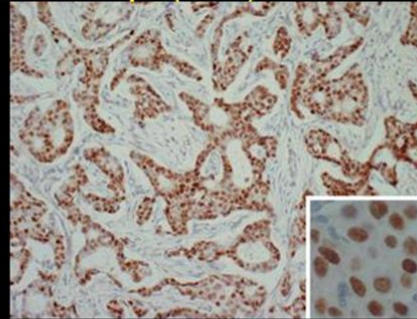
# More IHC Examples

## Oestrogen Receptor (ER): Run 65

IDC: 90 –95% Intensity: High  
Good example (above)  
Poor example (below)



IDC: 50 –75% Intensity: Medium  
Good example (above)  
Poor example (below)

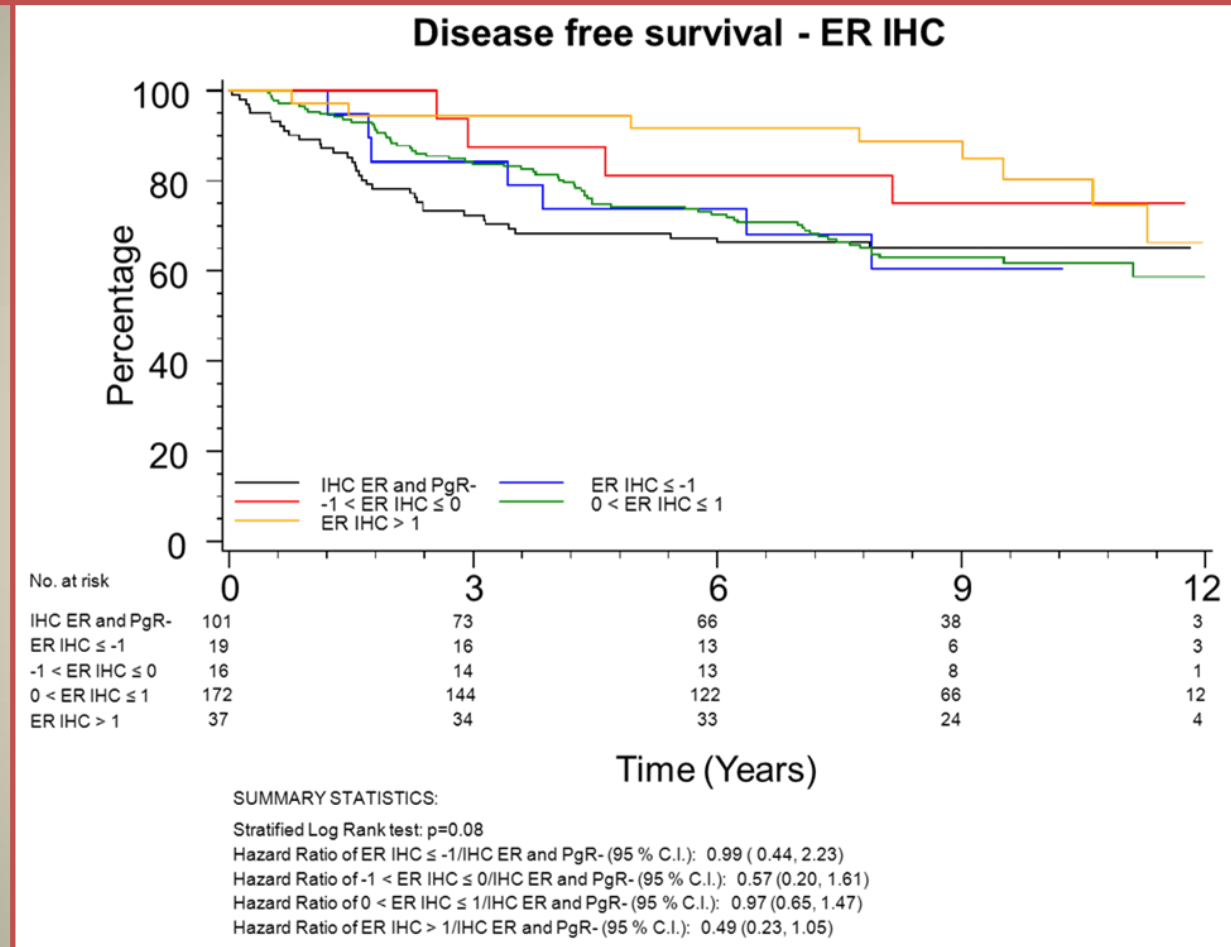


# Statistical Standardization

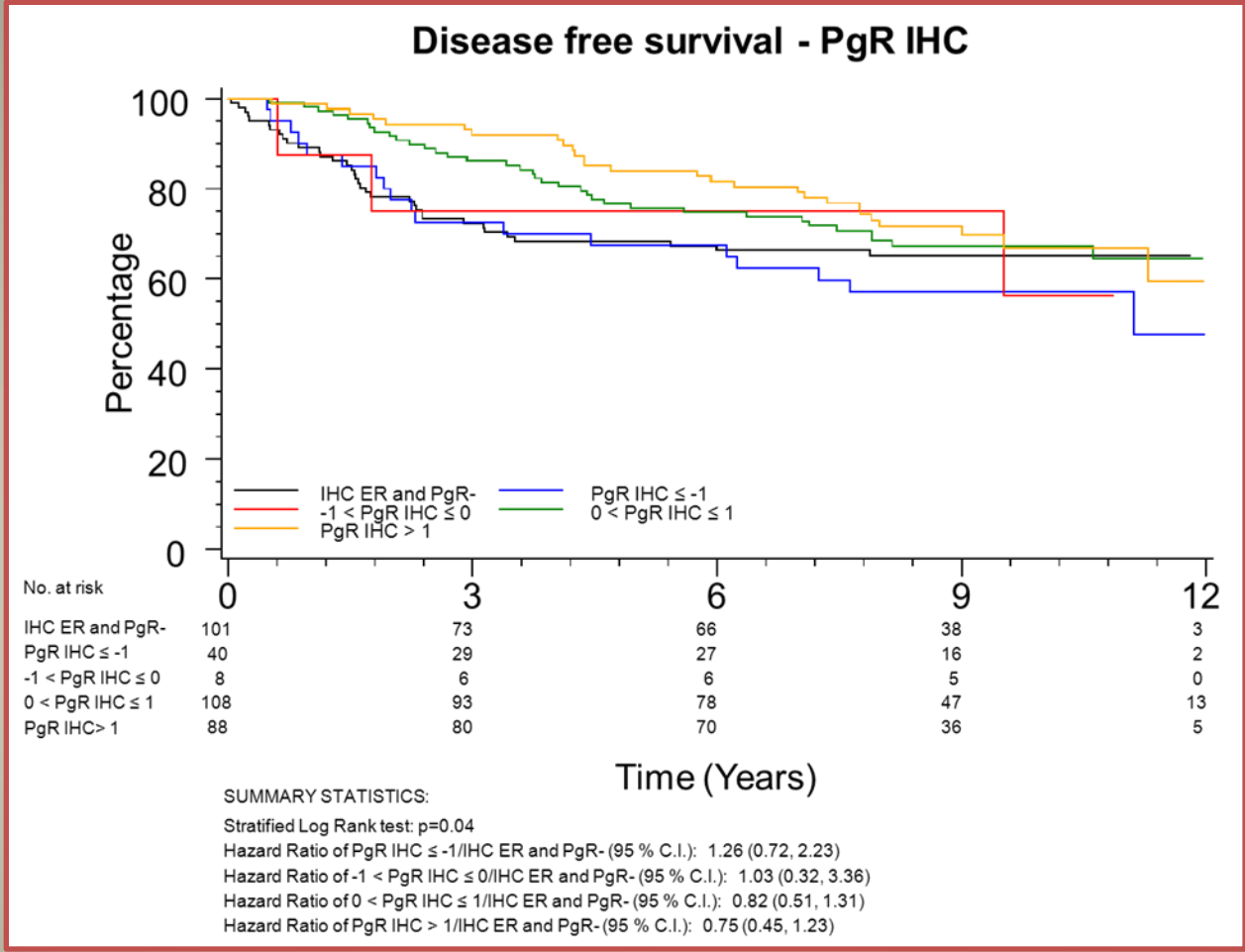
Akin to current World Health Organization mandated T-scores and Z-scores for Bone Mineral Density(BMD): work with standard deviations.



# NCIC CTG MA.12: both ER+/-

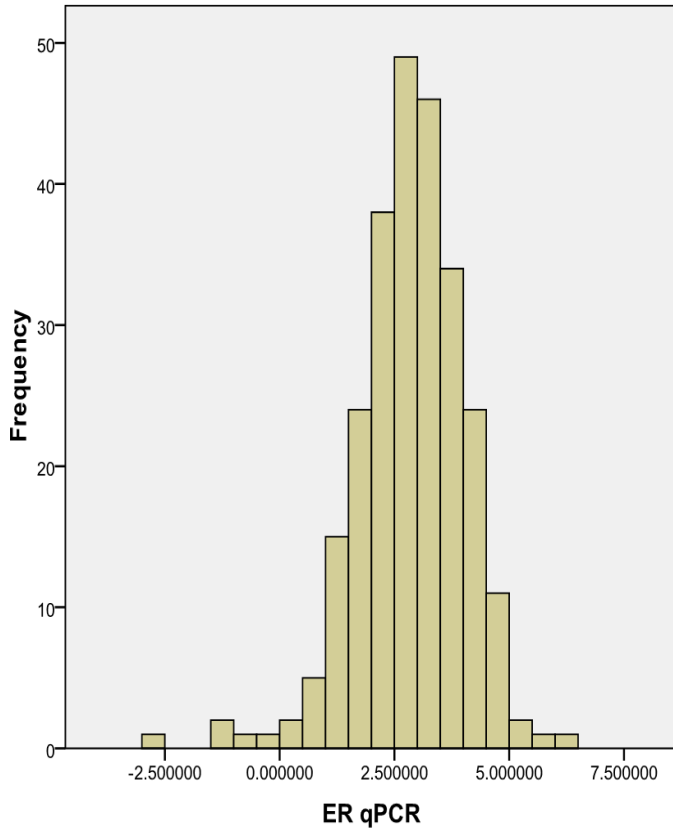


Chapman JW, Nielsen TO, et al. Breast Cancer Research 2013; 15:R71

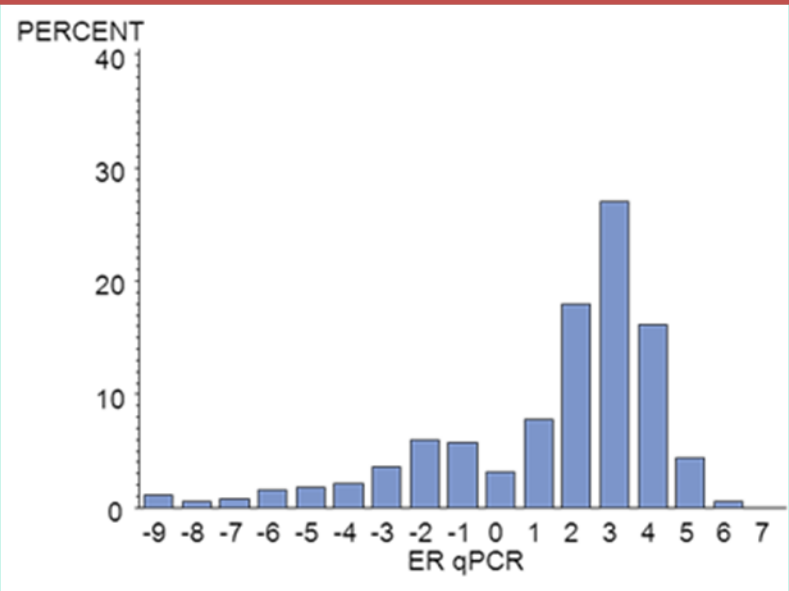
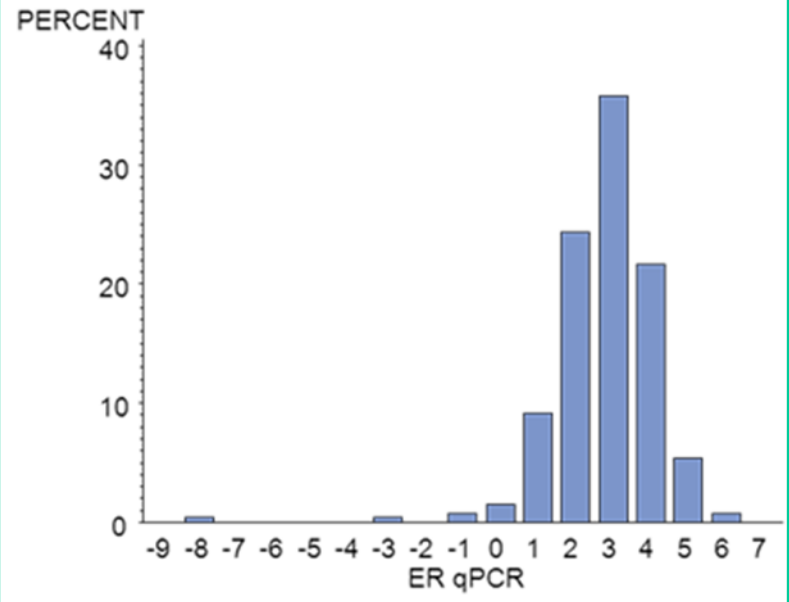


Chapman JW, Nielsen TO, et al. Breast Cancer Research 2013; 15:R71

MA.12 qPCR log(ER) among IHC ER+



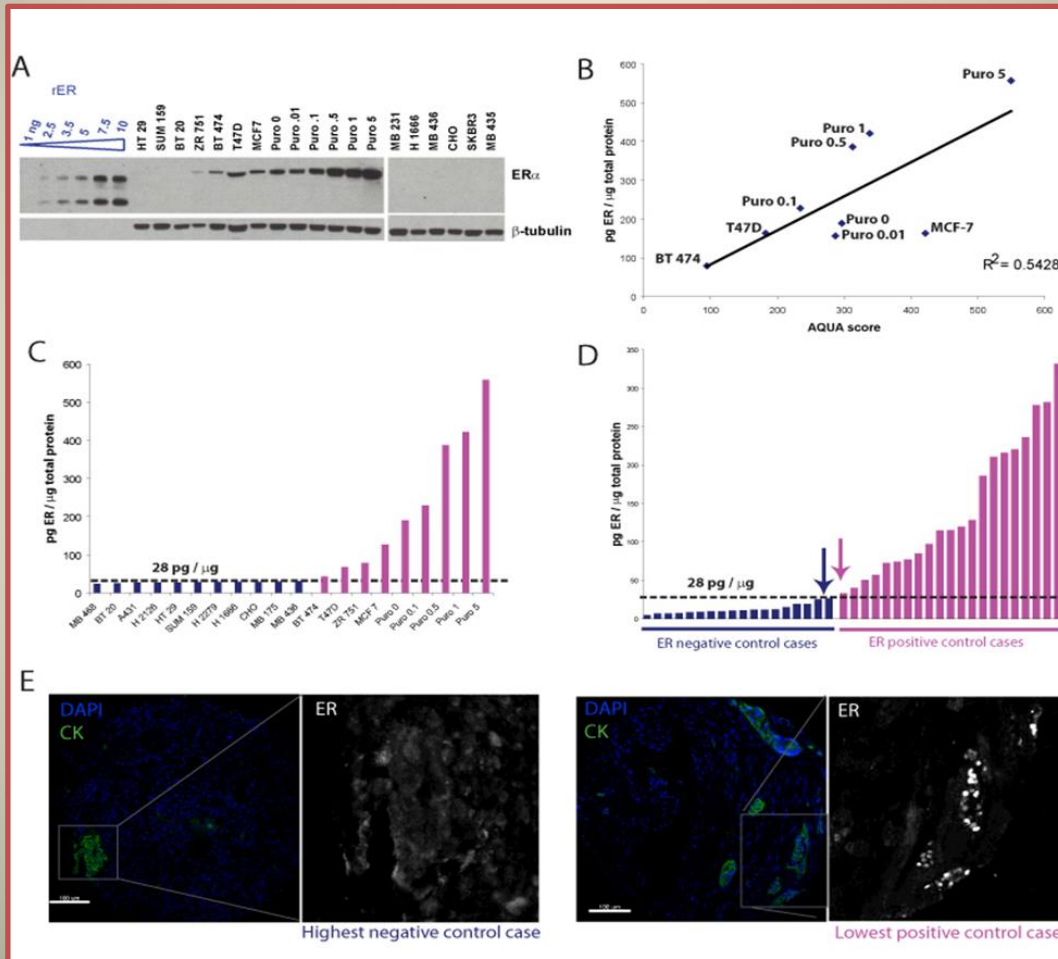
Mean =2.85  
Std. Dev. =1.186  
N=257



Chapman JW, Nielsen TO, et al. Breast Cancer Research 2013; 15:R71

JW Chapman

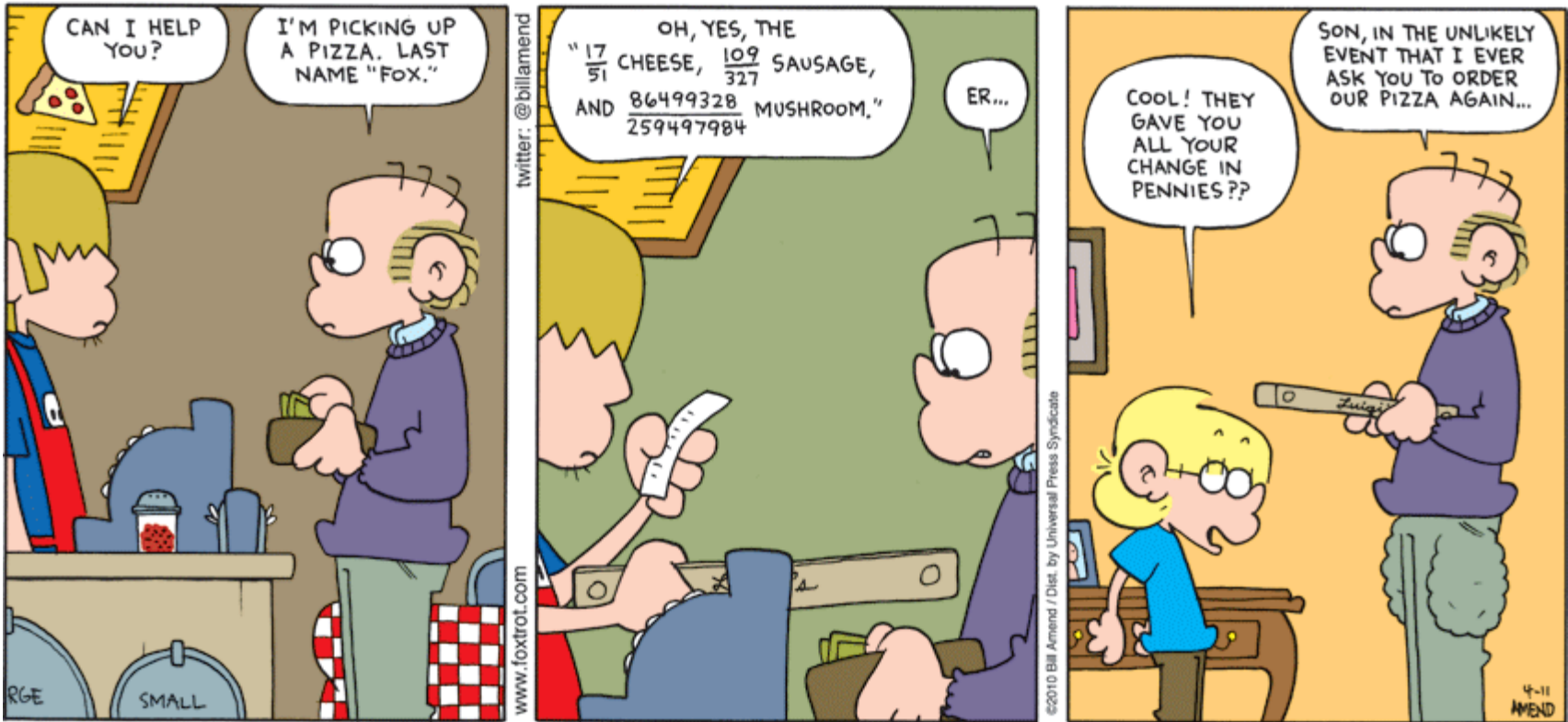
# David Rimm's AQUA



# Work in Progress

Statistical standardization of ER, PR, HER2, (ki67) using large NCIC CTG MA.27 trial (exemestane vs anastrozole) tumour samples, clinical follow-up: with Bharat Jasani, Keith Miller (UCL Advanced Diagnostics), Paul Goss and Lois Shepherd (MA.27), Sandip SenGupta (Queen's).

# Statistical details at appropriate level for area of practice?



# Communicated with good oral and written skills?



# Acknowledgements: mid 1980s →

- Betty Mobbs, Ontario Biochemical QC group; Betty Mobbs, Lavina Lickley, Kathy Pritchard, David McCready, Maureen Trudeau, Henrietta Banting Breast Centre (HBBC) group; David Murray, St. Michael's hospital (s-phase of HBBC tumours).
- Kananaskis Working Group, particularly Vince Shankey, Eric and Sandra Wolman.
- NCIC CTG MA.12: Torsten Nielsen, Vivien Bramwell, Lois Shepherd.
- NCIC CTG MA.14: Dennis Sgroi, Paul Goss, Michael Pollak, Lois Shepherd.
- UK NEQAS/UCL Advanced Diagnostics Bharat Jasani, Keith Miller; NCIC CTG MA.27 Paul Goss, Lois Shepherd; Sandip SenGupta (Queen's University).
- Patients who consented to use of their tumours and clinical follow-up for research purposes.